Hariprasad Vankayalapati and Gurdial Singh*

Department of Chemistry, University of Sunderland, Sunderland, UK SR1 3SD. E-mail: gurdial.singh@sunderland.ac.uk

Received (in Cambridge, UK) 6th April 2000, Accepted 2nd June 2000 Published on the Web 27th June 2000

The synthesis of the disaccharides α -L-Fucp-(1,2)- and α -L-Fucp-(1,3)- β -D-Galp(1)-4-methylumbelliferone as fucosidase substrates was accomplished by activation of the anomeric centre of 2,3,4-tri-O-acetyl- and 2,3,4,6-tetra-O-acetyl-α,β-L-fuco- and -D-galactopyranosyl propane-1,3-diyl phosphates with TMSOTf and with minimal protection of 4-methylumbelliferyl β-D-galactopyranoside.

Introduction

The importance of the role that carbohydrates play in the maintenance of health and the onset of disease has received due recognition during the last two decades. The diversity of these carbohydrate structures can be found in oligosaccharides that are covalently attached to lipids and proteins both at cell surfaces and in biological fluids. The length of these oligosaccharides is normally less than twenty sugar residues, yet the possible changes in configuration, oxidation and/or reduction states, and points of attachment give them a wide range of biological functions. Research in organic chemistry and molecular biology has been stimulated by the desire to attain an understanding of how and why these structures are recognised by enzymes, antibodies and lectins.

The fucose-containing disaccharides, e.g. L-Fuc- α -1,3- β -D-Galp and L-Fuc-α-1,2-β-D-Galp, are usually found at the non-reducing terminus of oligosaccharides of the blood-groupspecific glycoproteins and glycolipid with the structures L-Fuc-α-1,2-Gal-β-1,4-GlcNAc-β- and the grouping L-Fuc-α-1,3-Gal-β-1,4-Glu both occurring among human milk oligosaccharides.² Furthermore in sialyl Lewis X the α -Lfucopyranosyl group plays a pivotal role in binding to E-, L- and P-selectins.³ As part of our ongoing programme of synthesis of oligosaccharides that occur in human milk and of their analogues as fucosidase substrates, we describe the synthesis of both α -L-Fuc-1,3- β -galactopyranoses, α -L-Fuc-1,2- β galactopyranoses and also the corresponding disaccharides having a 4-methylumbelliferyl moiety attached at the anomeric centre of D-galactopyranose.

Results and discussion

DOI: 10.1039/b002754p

We chose to employ propane-1,3-diyl phosphate as the anomeric activating group for the synthesis of these compounds as we had successfully used it to obtain β-O-glycosides.⁴ In addition we reasoned that the displacement of this function with 4-methylumbelliferone would provide access to glycosides that are used as convenient substrates for the fluorimetric assay of glycoside hydrolyase activity⁵ and as ligands in carbohydrate protein interaction studies.⁶

Acetylation of L-fucopyranose 1 with acetic anhydride and pyridine, Scheme 1, afforded 1,2,3,4-tetra-O-acetyl-α,β-L-fucopyranose 2, which was converted to the α-L-fucopyranosyl bromide 3. Oxidation of the latter with silver carbonate in acetone gave the required 2,3,4-tri-O-acetyl-α,β-L-fucopyran-

$$H_3C$$
 OAC
 OAC

Scheme 1 Reagents and conditions: i, Ac₂O, Py, RT, 12 h; ii, 33% HBr-AcOH, RT, 15 min; iii, Ag₂CO₃, acetone, 0.5 h.

ose 4 in 90% overall yield. Treatment of 4 with propane-1,3dividioxyphosphoryl chloride 5^{4d} afforded the phosphates 6 and 7, as an inseparable crystalline mixture, in 65% yield, in the ratio 9:1 as determined by ¹H NMR. For the major isomer 6 a resonance at δ 5.92 was observed as a doublet of doublets (dd) for the anomeric proton and exhibited couplings of 3.3 and 2.6 Hz, whilst for the isomer 7 this resonance occurred at δ 5.35 with couplings of 5.9 and 7.9 Hz. The ¹³C NMR data were most informative for assignment of the stereochemistry at C-1 of 6 and 7; in the case for the former the resonance was observed at $\delta_{\rm C}$ 94.63 with a $J_{\rm C-P}$ of 4.9 Hz whilst for the latter anomer this was found at $\delta_{\rm C}$ 96.71 with $J_{\rm C-P}$ of

1,3,4,6-Tetra-O-acetyl-α-D-galactopyranose 8 was prepared as reported by Chittenden⁸ in a yield of 70%. Having both saccharides available we investigated the coupling of the phosphate 6 and 7 with 8, Scheme 2, and were gratified that this proceeded uneventfully when we employed trimethylsilyl triflate (TMSOTf) as the activating agent, and gave the peracetylated derivative of disaccharide α -L-Fucp-(1,2)- α -D-Galp, compound 9, in 63% yield after chromatographic purification on silica gel. The α -stereochemistry for the anomeric centre was assigned on

Scheme 2 Reagents and conditions: i, 1.5 equiv. TMSOTf, CH₂Cl₂, -78 °C; ii, NH₃(g), MeCN, 0 °C; iii, 5, N-MeIm, 16 h; iv, NaOMe, MeOH, RT, 30 min.

the basis of the observed ¹³C–¹H coupling constant of 165.4 Hz for the newly formed bond.9 To further establish that the coupling reaction had proceeded with the desired stereochemical outcome we undertook the removal of the acetate protecting groups in 9 with NaOMe in methanol and obtained the corresponding disaccharide which displayed spectral and physical properties that were in agreement with those reported by Lemieux.¹⁰ However, if the coupling of 6 and 7 with 8 was conducted with a catalytic amount of TMSOTf, the formation of the β-isomer of 9 was observed in 46% yield along with recovered starting material (39%). In this case a coupling of 159.6 Hz was observed for the 1,2-linkage. These observations suggest that the disaccharide 9 is the thermodynamic product and is the result of the bond-forming reaction proceeding via an oxonium ion intermediate whilst the corresponding β -isomer is formed via participation of the C-2 acetoxy function or by an $S_{\rm N}$ 2-like process.

The anomeric acetate group of 9 was selectively removed by employing ammonia gas in acetonitrile 11 and gave the disaccharide 10 in 69% yield after chromatographic purification. Treatment of 10 with propane-1,3-diyldioxyphosphoryl chloride 5 afforded the mixture of anomeric phosphates 11 in 60% yield, in the ratio 20:1 (11α major) as determined by ¹H NMR. In the ³¹P NMR spectrum the resonance for 11a occurred at δ_P –10.5 whilst that for 11β was found at δ_P –11.5. At this juncture we embarked upon a study of the displacement reaction of the mixture of phosphates 11 with 4-methylumbelliferone, as this function has been used extensively in the fluorimetric assay of glycosides. The propane-1,3-diyl phosphate group was successfully displaced by 4-methylumbelliferone using 0.2 mol equiv. of TMSOTf as the activator and afforded the β-linked disaccharide 12 in 56% yield following chromatography. This result is an improvement on the reported methods for the introduction of the 4-methylumbelliferone group which normally proceed in yields in the order of 14%. Removal of the acetate function of 12 yielded the fully deprotected disaccharide 13, Scheme 2. The stereochemical integrity of the anomeric centre of 13 was established on the basis of the ¹³C⁻¹H coupling constant which was determined as 159.2 Hz.

Having had success in the introduction of the 4-methylumbelliferone function we decided to investigate its introduction at the anomeric centre of galactose at an early stage and to study the subsequent coupling reaction with the fucopyranosyl phosphates 6 and 7, Scheme 3. Hence, selective removal of the anomeric acetate group of 148b afforded 2,3,4,6-tetra-O-acetylα,β-D-galactopyranose 15 in 96% yield. Treatment of 15 with propane-1,3-diyldioxyphosphoryl chloride 5 afforded the phosphates 16 as a mixture of separable crystalline α (mp 143– 145 °C) and β isomers in the ratio of 20:1, respectively, in a combined yield of 63%. The displacement of the phosphate 16α was accomplished with 4-methylumbelliferone and gave 4methylumbelliferyl† 2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranoside 17 in 68% yield; the same outcome was observed if the mixture of 16α,β was used. The stereochemistry of the anomeric linkage was assigned on the basis of a ¹³C-¹H coupling constant of 160.2 Hz. Subsequent deacetylation of the galactose derivative 17 with sodium methoxide in methanol cleanly afforded the saccharide 18 in 83% yield. The primary hydroxy group of the latter was protected as the silvl ether on treatment with tert-butyldiphenylsilyl chloride (TBDPSCl) and afforded the partially protected 4-methylumbelliferyl β-D-galactopyranoside 19 in 79% yield after chromatographic purification. At this juncture we investigated the chemistry of 19 with the fucopyranosyl phosphates 6 and 7, employing one mole equivalent of TMSOTf as the activator. This treatment resulted in the formation of the disaccharide 20 in 52% isolated yield along with a trace amount of the corresponding 1,2-linked disaccharide (23; see below) as evidenced by TLC; all of our attempts to isolate the 1,2-linked product by chromatography proved unfruitful. That the coupling reaction had proceeded regiospecifically at the C-3 OH function was tentatively assigned on the basis of ¹H and ¹³C NMR data. Removal of the primary silyl ether group afforded 21, which on treatment gave the fully

^{† &#}x27;4-Methylumbelliferyl' refers to 4-methylcoumarin-7-yl.

Scheme 3 Reagents and conditions: i, NH₃(gas), MeCN, 0 °C; ii, N-MeIm, CH₂Cl₂, 0 °C, 16 h; iii, TMSOTf, 0.2 equiv., -78 °C, 0.5 h; iv, NaOMe, MeOH, RT, 0.5 h; v, TBDPSCl, Im, DMF, RT, 8 h; vi, TMSOTf, 1 equiv., -78 °C, 1 h; vii, Bu₄NF, THF; viii, NaOMe, MeOH.

deprotected α -L-Fucp-1,3- β -D-Gal(1)-4-methylumbelliferone 22 in 59% yield for the two steps, Scheme 3. That the newly formed bond had a 1,3-linkage was strongly supported by the fact that the spectral and physical properties of 22 were different from those of the 1,2-linked isomer 13 that we had prepared earlier. In order to further support these conclusions we undertook the synthesis of both 13 and 22 by employing classical coupling chemistry, Scheme 4. Thus coupling of 2,3,4-tri-O-acetyl-α-Lfucopyranosyl bromide 3 with the partially protected derivative 19 using tetraethylammonium bromide afforded the disaccharides 20 and 23 in yields of 27% and 51%, respectively. Removal of the silyl and acetate functions resulted in the formation of the isomeric fully deprotected disaccharides 22 and 13 which had spectral and physical properties in agreement with the compounds prepared using the propane-1,3-diyl phosphate coupling protocol, Schemes 2 and 3.

Conclusions

In summary we have established that fucose-containing disaccharides can be prepared using propane-1,3-diyl phosphate activation of the anomeric centre of glycosides in a regio/stereoselective manner.

Experimental

General methods

¹H NMR spectra were recorded in CDCl₃ (unless stated otherwise) on a JEOL GSX 270 NMR spectrometer and are reported

as follows: chemical shifts δ (ppm), [number of protons, multiplicity, coupling constants J (Hz), and assignment]. Residual protic solvent CHCl₃ ($\delta_{\rm H}$ 7.26) was used as the internal reference. ¹³C NMR spectra were recorded in CDCl₃, at 67.8 MHz on the JEOL GSX 270 NMR spectrometer, using the central resonance of CDCl₃ ($\delta_{\rm C}$ 77.0) as the internal reference. ¹³P NMR spectra were recorded in CDCl₃, at 109.25 MHz on a JEOL GSX 270 NMR spectrometer, using trimethyl phosphate as the external reference. IR spectra were recorded on a UNICAM series FTIR spectrometer. Mass spectra were obtained on an AEI MS 902 or a VG ZAB-E spectrometer. UVvisible spectra were measured on a UNICAM UV/VIS spectrophotometer. Microanalysis were performed by MEDAC Ltd, Surrey. Mps were determined on a GallenKamp capillary melting-point apparatus and are uncorrected. Optical rotations were measured in CHCl₃ solution using a Bellingham & Stanley ADP 220 polarimeter. $[a]_D$ -Values are in units of 10^{-1} deg cm² g⁻¹. Flash chromatography was carried out using Fluka silica gel 60 (230-400 mesh) unless otherwise indicated. Analytical TLC was performed using precoated aluminium plates (Merck Kiselgel 60 F₂₅₄) and visualised by UV, phosphomolybdic acid or basic aq. potassium permanganate solutions. 'Petrol' refers to petroleum spirit (distillation range 40-60 °C) which was distilled prior to use, and ether refers to diethyl ether.

All reactions were carried out under an argon or nitrogen atmosphere in oven-dried glassware unless otherwise stated. CH₂Cl₂ and DMF were distilled from calcium hydride and stored over 4 Å molecular sieves. Aqueous solutions are saturated unless otherwise specified.

Scheme 4 Reagents and conditions: ii, Bu₄NF, THF; iii, NaOMe, MeOH, RT, 0.5 h.

1,2,3,4-Tetra-O-acetyl- α , β -L-fucopyranose 2⁷

A solution of L-fucose 1 (5 g, 30.45 mmol) in dry pyridine (50 ml) containing acetic anhydride (35 ml) was stored at 4 °C for 12 h. Following this water (10 ml) was added and, after 1 h, the solution was extracted with CH_2Cl_2 (100 ml). The combined CH_2Cl_2 layers were back-extracted with 100 ml of cold 1 M H_2SO_4 and then with cold aq. NaHCO₃ (100 ml), and finally washed with water (100 ml). The organic phase was dried (Na₂SO₄) and concentrated to give α,β -L-fucopyranosyl tetraacetate 2 as a thick syrup (11.00 g, 100%).

For isomer **2a**, IR (film) $v_{\rm max}$ 1751, 1371 cm⁻¹; $\delta_{\rm H}$ 1.29 (3H, d, J 6.6), 2.00 (3H, s), 2.05 (3H, s), 2.10 (3H, s), 2.18 (3H, s), 4.08 (1H, appt d, J 7.3), 4.39 (1H, appt d, J 6.6), 5.10–5.16 (1H, m), 5.39 (1H, d, J 3.3), 6.29 (1H, d, $J_{1,2}$ 4.6, H-1); $\delta_{\rm C}$ 15.97, 20.44, 20.56, 20.60, 20.64, 67.65, 69.04, 70.19, 83.22, 90.29 (C-1), 169.88, 170.12 (2C), 170.47.

For isomer **2β**, $\delta_{\rm H}$ 1.13 (3H, d, J 6.6), 2.09 (3H, s), 2.12 (3H, s), 2.13 (3H, s), 2.19 (3H, s), 3.86–3.92 (1H, m), 4.20–4.34 (1H, m), 5.01–5.07 (1H, m), 5.30 (1H, dd, J 3.3, 1.3), 5.43 (1H, d, $J_{1,2}$ 7.3, H-1); $\delta_{\rm C}$ 15.91, 20.41, 20.49, 20.57, 20.90, 64.02, 68.39, 70.84, 82.07, 95.45 (C-1), 170.01, 170.32, 170.65, 170.82.

2,3,4-Tri-O-acetyl- α -L-fucopyranosyl bromide 3 7

The mixture of α,β-fucopyranosyl tetraacetates **2** (10 g, 30.1 mmol) was dissolved in glacial acetic acid (20 ml) and 30 ml of 33% HBr–HOAc was added slowly at 20 °C. Stirring was continued for 15 min and the yellow coloured solution was dissolved in CH₂Cl₂ (200 ml), and back-extracted with cold water (2 × 200 ml), cold 1 M aq. NaHCO₃ (200 ml) and finally with 200 ml of water. The CH₂Cl₂ layer was dried over Na₂SO₄ and concentrated *in vacuo* to give the α-L-fucopyranosyl bromide **3** as a thick syrup (10.32 g, 97%), $\delta_{\rm H}$ 1.21 (3H, d, J 7.3), 2.05 (3H, s), 2.11 (3H, s), 2.18 (3H, s), 4.08 (1H, appt d, J 7.3), 4.37 (1H, appt d, J 6.6), 5.00 (1H, appt d, J 4.0), 5.36 (1H, dd, J 3.3, 2.6), 6.68 (1H, d, J_{1,2} 4.0, H-1); $\delta_{\rm C}$ 15.50, 20.49, 20.56, 20.72, 67.84, 68.40, 69.79, 69.98, 89.27, 169.79, 170.12, 170.25.

2,3,4-Tri-*O*-acetyl-α,β-L-fucopyranose 4⁷

The α -L-fucopyranosyl bromide 3 (9.5 g, 26.9 mmol) was dissolved in acetone (50 ml) containing 1.4 ml of water at 0 °C. To the resulting solution was added Ag₂CO₃ (7.4 g, 26.9 mmol) and the mixture was stirred for 0.5 h. The reaction mixture was filtered through a pad of Celite and the filtrate was evaporated to give a thick syrup, which was purified by column chromato-

graphy (1:1 EtOAc–petrol) and afforded 2,3,4-tri-O-acetyl- α , β -L-fucopyranose 4 (7.35 g, 94%).

For isomer 4α , IR (film) $\nu_{\rm max}$ 3357, 1745, 1369 cm⁻¹; $\delta_{\rm H}$ 1.24 (3H, d, J 6.6), 2.00 (3H, s), 2.10 (3H, s), 2.17 (3H, s), 4.39 (1H, appt d, J 6.6), 5.12 (1H, dd, J 4.0, 3.3), 5.31 (1H, dd, J 4.0, 1.3), 5.43 (1H, dd, J 4.0, 3.3), 5.46 (1H, d, $J_{1,2}$ 3.3, H-1); $\delta_{\rm C}$ 15.86, 20.55, 20.60, 20.75, 64.33, 67.72, 68.45, 71.26, 90.57 (C-1), 170.13, 170.43, 170.66.

For isomer **4β**, $\delta_{\rm H}$ 1.14 (3H, d, J 6.6), 2.05 (3H, s), 2.10 (3H, s), 2.18 (3H, s), 3.83 (1H, appt d, J 6.6), 4.64 (1H, d, $J_{1,2}$ 7.9, H-1), 5.01 (1H, dd, J 5.3, 2.6), 5.25 (1H, d, J 1.3), 5.39 (1H, d, J 4.0); $\delta_{\rm C}$ 14.09, 20.50, 20.53, 20.93, 60.38, 69.42, 70.26, 70.78, 95.75 (C-1), 170.10, 170.47, 170.57.

2', 3', 4' -Tri- ${\it O}$ -acetyl- α, β -L-fucopyranosyloxy-1,3,2-dioxaphosphinane 2-oxide 6 and 7

2,3,4-Tri-O-acetyl- α , β -L-fucopyranose 4 (5.00 g, 17.2 mmol) was dissolved in CH₂Cl₂ (50 ml) and the solution was cooled to 0 °C. To this solution were added 2-chloro-1,3,2-dioxaphosphinane 2-oxide (5.4 g, 34.39 mmol) and 1-methylimidazole (2.8 g, 34.48 mmol) dropwise over a period of 15 min. Stirring was continued for 16 h, with gradual warming to room temperature. The solvent was removed, the residue was re-dissolved in CH₂Cl₂ (50 ml), and the solution was evaporated to remove traces of 1-methylimidazole. Addition of CH₂Cl₂ (50 ml) to the residue gave a pale yellow solution, which was washed successively with ice-water (50 ml), aq. NaHCO₃ (2 × 50 ml) and water (50 ml). The organic layer was dried over Na₂SO₄ and the solvent was removed in vacuo. The oily residue was purified by column chromatography (EtOAc-petrol 1:1) to afford the phosphates 6 and 7 (ratio 9:1) as an inseparable crystalline mixture (4.68 g, 65%), mp_{mix} 151–153 °C; $[a]_D^{22}$ –100.6 (c 3.2, CHCl₃).

For isomer **6**, IR (KBr) $v_{\rm max}$ 1745, 1373, 1216 cm⁻¹; $\delta_{\rm H}$ 1.24 (3H, d, J 6.6, C H_3), 1.85–1.89 (1H, m, $J_{\rm P-H}$ 15.17, H^{ax}-5), 2.02 (3H, s), 2.10 (3H, s), 2.19 (3H, s), 2.30–2.36 (1H, m, $J_{\rm P-H}$ 15.2, H^{eq}-5), 3.90 (1H, dq, J 7.3, 6.6), 4.33–4.59 (4H, m), 5.01 (1H, appt d, J 3.3, 3.3), 5.22 (1H, dd, J 3.3, 2.6), 5.31–5.33 (1H, m), 5.92 (1H, dd, $J_{1,2}$ 3.3, 2.6, H-1); $\delta_{\rm C}$ 15.82, 20.52, 20.60 (2C), 25.63, 66.92, 67.35, 68.80, 68.86 (2C), 69.75, 70.36, 94.63 (d, $J_{\rm C-P}$ 4.93, C-1), 170.14 (2C), 170.46; $\delta_{\rm P}$ –10.36.

For isomer 7 $\delta_{\rm H}$ 1.19 (3H, d, J 6.6), 1.78–1.83 (1H, m, $J_{\rm P-H}$ 15.2, H^{ax}-5), 2.02 (3H, s), 2.10 (3H, s), 2.19 (3H, s), 2.23–2.28 (1H, m, $J_{\rm P-H}$ 15.2, H^{eq}-5), 3.67 (1H, dq, J 6.0, 5.9), 4.07–4.31 (2H, m), 4.33–4.59 (4H, m), 5.31–5.33 (1H, m), 5.35 (1H, dd,

 $J_{1,2}$ 5.9, 7.9, H-1); $\delta_{\rm C}$ 15.85, 20.49, 20.65, 20.74, 25.79, 66.83, 68.91, 68.97, 69.31, 70.27, 70.74, 96.71 (d, $J_{\rm C-P}$ 4.41, C-1), 170.41, 170.43, 170.49; $\delta_{\rm P}$ –10.80; m/z (CI, NH₃) (Found: M⁺, 410.0978. C₁₅H₂₃O₁₁P requires M, 410.0978) (Found: C, 43.68; H, 5.58; P, 7.58. C₁₅H₂₃O₁₁P requires C, 43.91; H, 5.65; P, 7.55%).

1,3,4,6-Tetra-O-acetyl-α-D-galactopyranose 8

The title compound was prepared as detailed by Chittenden.⁸ Thus β-D-galactose pentaacetate (5 g, 12.8 mmol) in TFA (17.5 ml) containing water (1.75 ml) was stirred at RT for 5 h and concentrated *in vacuo*. The residue was dissolved in toluene (20 ml), which was subsequently removed *in vacuo*. The residual product was crystallised from Prⁱ₂O to give **8** (6.23 g, 70%), mp 149–151 °C; $[a]_D^{10}$ +125.3 (*c* 1.4, CHCl₃); IR (film) v_{max} 3444, 1747, 1373 cm⁻¹; δ_H 2.04 (3H, s), 2.06 (3H, s), 2.15 (3H, s), 2.18 (3H, s), 4.07–4.09 (2H, m), 4.14 (1H, dd, *J* 10.6, 4.0), 4.26 (1H, t, *J* 6.6), 5.15 (1H, appt d, *J* 3.3), 5.44 (1H, dd, *J* 3.3, 1.3), 6.29 (1H, d, $J_{1,2}$ 4.0, H-1); δ_C 20.49, 20.55, 20.67, 20.86, 61.23, 65.87, 67.48, 68.60, 70.27, 91.87 (C-1), 169.49, 170.08, 170.44, 170.88.

1',3',4',6'-Tetra-*O*-acetyl-2'-*O*-(2,3,4-tri-*O*-acetyl-α-L-fuco-pyranosyl)-α-D-galactopyranose 9

Propane-1,3-diyl 2,3,4-tri-O-acetyl-α,β-L-fucopyranosyl phosphate **6/7** (2.0 g, 4.9 mmol) was dissolved in dry CH₂Cl₂ (20 ml) and the solution was cooled to -78 °C. To the resultant solution was added TMSOTf (1.62 g, 7.3 mmol) and, after 5 min stirring, a solution of glycosyl acceptor 8 (1.69 g, 4.85 mmol) in CH₂Cl₂ (5 ml) was added. The reaction mixture was stirred at -78 °C for 30 min and was then allowed to warm up to 0 °C. The reaction mixture was quenched with aq. NaHCO₃ (20 ml) and extracted into CH₂Cl₂ (20 ml). The organic layer was dried over anhydrous Na₂CO₃, concentrated, and the residue was purified by column chromatography (EtOAc-'petrol' 1:1) to give compound **9** (1.93 g, 63%), mp 113–117 °C; $[a]_D^{22}$ +32.85 (c 2.8, CHCl₃); IR (film) v_{max} 1751, 1373 cm⁻¹; δ_{H} 1.19 (3H, d, J 6.6), 1.97 (3H, s), 2.00 (3H, s), 2.02 (3H, s), 2.03 (3H, s), 2.04 (3H, s), 2.14 (3H, s), 2.17 (3H, s), 3.78 (1H, dd, *J* 6.6, 5.9), 4.06– 4.16 (3H, m), 4.25 (1H, t, J 6.6), 4.49 (1H, d, J 7.9), 4.94 (1H, dd, J 9.9, 3.3), 5.07 (1H, dd, J 10.6, 7.9), 5.20 (1H, d, J 2.6), 5.28 (1H, dd, J 9.2, 3.3), 5.47 (1H, d, $J_{1,2}$ 2.6, H-1), 6.26 (1H, d, $J_{1',2'}$ 4.0, H-1); $\delta_{\rm C}$ 16.12 (CH₃), 20.49, 20.57 (3C), 20.65 (2C), 20.70, 61.09, 67.73, 68.09, 68.38, 68.69, 69.22, 69.90, 71.34, 73.76, 89.77 (C-1'), 101.67 (C-1), 169.22, 169.33, 169.84, 169.94, 170.12, 170.35, 170.58; m/z (CI, NH₃) [Found: (M + NH₄), 638.2294. $C_{26}H_{40}NO_{17}$ requires m/z, 638.2296].

3,4,6-Tri-O-acetyl-2-O-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)- α , β -D-galactopyranose 10

To solution of ammonia in CH₃CN (25 ml), prepared by bubbling ammonia gas through the solvent at 0 °C (10 min), was added compound **9** (1.5 g, 2.4 mmol) and the mixture was stirred at RT for 24 h. The solvent was then removed *in vacuo* and the residue was purified by column chromatography (EtOAc-'petrol' 1:1) to give a mixture of **10** α and **10** β anomers in the ratio 3:1 (960 mg, 69%), mp 74–78 °C; [a]_D¹⁹ +24.6 (c 2.4, CHCl₃).

For isomer 10a IR (KBr) $v_{\rm max}$ 3352, 1753 cm⁻¹; $\delta_{\rm H}$ 1.24 (3H, t, J 7.3), 1.99 (3H, s), 2.01 (3H, s), 2.03 (3H, s), 2.05 (3H, s), 2.07 (3H, s), 2.18 (3H, s), 3.69 (1H, dd, J 7.9, 2.0), 3.79 (1H, t, J 7.9), 3.88 (1H, m), 3.98–4.20 (2H, m), 4.66 (1H, d, J 7.9), 4.72 (1H, dd, J 7.9, 4.6), 5.00 (1H, dd, J 7.3, 3.3), 5.08–5.18 (2H, m), 5.21 (1H, d, $J_{1',2'}$ 3.3), 5.30–5.37 (1H, m), 5.39 (1H, d, $J_{1,2}$ 3.3); $\delta_{\rm C}$ 16.02 (C H₃), 20.51, 20.57 (2C), 20.66, 20.70, 20.94, 60.34, 61.46, 67.38, 68.14, 69.05, 69.45, 70.00, 71.03, 71.46, 91.53 (C-1'), 96.22 (C-1), 169.70, 169.81, 170.09, 170.14, 170.61, 170.52.

For isomer **10** β , (selected features) $\delta_{\rm H}$ 1.18 (3H, t, J 5.94, C H_3), 4.44 (1H, d, $J_{1,2}$ 6.6, H-1); $\delta_{\rm C}$ 101.86 (C-1).

2-[3',4',6'-Tri-*O*-acetyl-2'-*O*-(2",3",4"-tri-*O*-acetyl- α -L-fucopyranosyl)- α , β -D-galactopyranosyloxy]-,1,3,2-dioxaphosphinane 2-oxide 11 α and 11 β

The title compounds were prepared using a similar procedure to that employed for compounds **6** and **7**, by treating compound **10** (1.2 g, 2.1 mmol) with propane-1,3-diyldioxyphosphoryl chloride **5** (0.64 g, 4.0 mmol) in CH₂Cl₂ (15 ml) and 1-methylimidazole (0.34 g, 4.15 mmol) at RT for 16 h. Chromatographic purification of the resulting residue (EtOAc-'petrol' 1:1) gave an inseparable crystalline mixture of **11** α and **11** β (ratio 9:1) (861 mg, 60%), mp 109–112 °C; [a]²⁰ +36.23 (c 2.8, CHCl₃).

For isomer 11 α , IR (KBr) $\nu_{\rm max}$ 1749, 1214 cm⁻¹; $\delta_{\rm H}$ 1.23 (3H, d, J 7.3), 1.78–1.84 (1H, m, $J_{\rm P-H}$ 15.2, Hax-5), 2.24–2.30 (1H, m, $J_{\rm P-H}$ 15.2, Heq-5), 1.97 (3H, s), 2.04 (3H, s), 2.05 (6H, s), 2.08 (3H, s), 2.17 (3H, s), 3.80 (1H, appt d, J 8.6), 3.92–4.04 (2H, m), 4.07 (1H, dd, J 7.3, 3.3), 4.15 (1H, t, J 5.3), 4.43–4.54 (4H, m), 4.62 (1H, d, J 7.9), 4.96 (1H, dd, J 9.2, 2.0), 5.07 (1H, d, J 7.3), 5.21–5.38 (2H, m), 5.43 (1H, d, $J_{1',2'}$ 2.6, H-1"), 5.90 (1H, dd, $J_{1',2'}$ 3.3, H-1'); $\delta_{\rm C}$ 16.00 (C H₃), 20.33, 20.38, 20.42, 20.51, 20.60, 20.79, 25.62, 61.67, 66.64, 67.39, 67.65, 68.65, 68.45 (d, J 4.67), 68.79, 68.94, 69.26, 69.97, 71.58, 95.14 (d, J 5.46, C-1'), 95.39 (d, J 6.23, C-1"), 169.15, 169.68, 169.87, 169.98, 170.21, 171.41; $\delta_{\rm P}$ -10.54.

For isomer 11 β , (selected features) $\delta_{\rm H}$ 1.20 (3H, d, J 6.6), 5.82 (1H, dd, $J_{1',2'}$ 6.6, H-1'); $\delta_{\rm C}$ 97.88 (d, J 5.71, C-1"); $\delta_{\rm P}$ -11.50; m/z (CI, NH₃) [Found: (M + NH₄), 716.1266. C₂₇H₄₃NO₁₉P requires m/z, 716.1267] (Found: C, 46.46; H, 5.52; P, 4.65. C₂₇H₃₉O₁₉P requires C, 46.42; H, 5.58; P, 4.44%).

4-Methylumbelliferyl 3',4',6'-tri-*O*-acetyl-2'-(2",3",4"-tri-*O*-acetyl-α-L-fucopyranosyl)-β-D-galactopyranoside 12

The title compound was prepared using a similar method to that employed for 9, by activating phosphates $11\alpha\beta$ (1 g, 1.4 mmol) with TMSOTf (0.08 g, 0.35 mmol) at -78 °C in CH₂Cl₂ (10 ml). Following the addition of 4-methylumbelliferone (0.25 g, 1.43 mmol) at the same temperature the reaction mixture was stirred for 0.5 h and allowed to warm to 0 °C. The solution was quenched with aq. NaHCO₃ (10 ml) and the solvent was removed in vacuo. The resultant residue was dissolved in CH₂Cl₂ (10 ml), and the extract was dried over Na₂CO₃, concentrated, and purified by column chromatography (EtOAc-'petrol' 1:1) to afford **12** as a white solid (560 mg, 56%), mp 121–123 °C; $[a]_D^{18}$ +37.2 (c 0.8, CHCl₃); IR (KBr) v_{max} 1745, 1614, 1371 cm⁻¹; $\delta_{\rm H}$ 1.15 (3H, d, J 6.6), 1.96 (3H, s), 1.98 (3H, s), 2.03 (3H, s), 2.05 (3H, s), 2.07 (3H, s), 2.13 (3H, s), 2.43 (3H, s, CH₃), 3.93 (1H, dd, J 7.3, 6.6), 4.07–4.16 (3H, m), 4.24 (1H, dd, J 5.3, 4.6), 4.56 (1H, appt d, J 7.9), 4.95 (1H, dd, J 6.6, 4.0), 5.15 (1H, d, $J_{1'',2''}$ 7.9, H-1"), 5.19 (1H, dd, J 7.3, 4.0), 5.34 (1H, d, J 3.3), 5.47–5.52 (1H, m), 5.67 (1H, dd, $J_{1',2'}$ 4.0, 3.3, H-1'), 6.20 (1H, s), 7.02 (1H, dd, J 7.9, 2.0), 7.12 (1H, d, J 4.0), 7.53 (1H, d, J 9.2); $\delta_{\rm C}$ 16.00 (CH₃), 18.59 (CH₃), 20.47, 20.47, 20.50, 20.57, 20.60, 20.62, 20.96, 61.25, 67.58, 67.86, 68.44, 69.46, 69.91, 71.20, 72.95, 83.26, 96.04 (C-1'), 103.85 (C-1"), 104.24, 113.04, 114.14, 115.20, 125.74, 152.18, 154.84, 159.12, 168.83, 169.86, 170.04, 170.08, 170.28, 170.52, 170.54; m/z (CI, NH₃) [Found: $(M + NH_4)$, 754.2560. $C_{34}H_{44}NO_{18}$ requires m/z, 754.2558].

4-Methylumbelliferyl 2'-O-(α -L-fucopyranosyl)- β -D-galactopyranoside 13

To a solution of the disaccharide 12 (500 mg, 0.68 mmol) in methanol (50 ml) was added NaOMe (0.055 g, 1 mmol) and the resultant solution was stirred at RT for 0.5 h. The solution was neutralised by passage down a Dowex 50 (H⁺) ion-exchange resin. Evaporation of the combined fractions gave the title compound as a white solid (205 mg, 62%), mp 251–253 °C; $[a]_D^{12} + 24.7$ (c 0.7, H₂O); IR (KBr) v_{max} 3480, 1709, 1615, 834 cm⁻¹;

 $\begin{array}{l} \lambda_{\rm max} \left({\rm H_2O} \right) 318, 294, 252 \ {\rm nm}; \\ \delta_{\rm H} \left({\rm DMSO\text{-}d_6} \right) 1.09 \left({\rm 3H, d, } J \, 6.6 \right), \\ 2.39 \left({\rm 3H, s} \right), 3.46\text{--}3.69 \left({\rm 3H, m} \right), 3.53 \left({\rm 1H, appt d, } J \, 6.0 \right), 3.76 \\ \left({\rm 1H, dd, } J \, 10.0, \, 7.3 \right), 3.86 \left({\rm 1H, br d, } J \, 4.0 \right), 3.92 \left({\rm 1H, br d, } J \, 2.7 \right), \\ 4.06 \left({\rm 1H, dd, } J \, 9.6, 3.3 \right), 4.24 \left({\rm 1H, q, } J \, 7.3 \right), \\ 4.34 \left({\rm 1H, dd, } J \, 5.3, \, 3.3 \right), 5.09 \left({\rm 1H, appt d, } J_{1',2'} \, 8.0, \, {\rm H\text{--}}1'' \right), \\ 5.70 \left({\rm 1H, d, } J_{1',2'} \, 3.30, \, {\rm H\text{--}}1' \right), \\ 6.23 \left({\rm 1H, s} \right), 7.03 \left({\rm 1H, dd, } J \, 7.3, \, 2.0 \right), \\ 7.09 \left({\rm 1H, d, } J \, 2.7 \right), \\ 7.67 \left({\rm 1H, d, } J \, 9.3 \right); \\ \delta_{\rm C} \left({\rm DMSO\text{--}d_6} \right) \, 16.52, \\ 18.10, 60.21, \\ 67.82, 68.18, 69.06, 70.62, 71.00, 72.53, 73.12, 73.21, 96.91 \left(J_{\rm CH} \, 173.5, \, {\rm C\text{--}}1'' \right), \\ 101.99 \left(J_{\rm CH} \, 159.2, \, {\rm C\text{--}}1' \right), \\ 104.16, \\ 111.73, \\ 114.21, \\ 126.36, \\ 153.33, \\ 154.32, \\ 159.99, \\ 160.11; \\ m/z \left({\rm CI, NH_3} \right) \\ [{\rm Found:} \left({\rm M + NH} \right), \\ 502.1925, \\ {\rm C_{22}H_{37}NO_{12}} \ {\rm requires} \, 502.1924 \right]. \\ \end{array}$

2,3,4,6-Tetra-O-acetyl-α,β-D-galactopyranose 15

β-D-Galactose pentaacetate **14** (5 g, 12.8 mmol) was added to a solution of ammonia in CH₃CN (200 ml), prepared by bubbling ammonia gas through the solvent at 0 °C (20 min). The mixture was stirred at RT for 24 h. The solvent was removed *in vacuo* and the residue was purified by column chromatography (EtOAc-'petrol' 3:2) to afford tetraacetate **15** (4.32 g, 96%), $[a]_D^{19} + 31.66$ (c 1.2, CHCl₃).

For α isomer, IR (film) $v_{\rm max}$ 3417, 1747, 1371 cm $^{-1}$; $\delta_{\rm H}$ 2.07 (3H, s), 2.16 (6H, s), 2.18 (3H, s), 4.13–4.18 (2H, m), 4.21 (1H, dd, J 7.9, 6.6), 5.07 (1H, d, J 7.9), 5.34 (1H, d, J 4.0), 5.50 (1H, d, J 3.3), 6.32 (1H, d, $J_{1,2}$ 3.3, H-1); $\delta_{\rm C}$ 20.52, 20.55, 20.59, 20.70, 61.73, 66.06, 67.23, 68.17, 68.35, 90.55 (C-1), 170.10, 170.28, 170.43, 170.60.

For β isomer, $\delta_{\rm H}$ 1.98 (3H, s), 2.00 (3H, s), 2.03 (3H, s), 2.04 (3H, s), 3.90 (1H, dd, J 7.3, 6.6), 4.05–4.11 (2H, m), 4.57 (1H, d, J 1.2, 7.9, H-1), 4.93 (1H, q, J 3.3), 5.29 (1H, d, J 3.3), 5.43 (1H, d, J 2.64); $\delta_{\rm C}$ 20.44, 20.49, 20.57, 20.65, 60.40, 61.41, 67.12, 70.41, 70.91, 95.85 (C-1), 170.06, 170.20, 170.55, 170.95.

2-(2',3',4',6'-Tetra-O-acetyl- α,β -D-galactopyranosyloxy)-1,3,2-dioxaphosphinane 2-oxide 16α and 16β

Treatment of 2,3,4,6-tetra-O-acetyl- α ,β-D-galactopyranose 15 (5 g, 14.4 mmol) with propane-1,3-diyldioxyphosphoryl chloride (4.5 g, 29.0 mmol) in CH₂Cl₂ (50 ml) and 1-methylimidazole (2.35 g, 28.7 mmol), at RT for 16 h as for the preparation of 6 above, afforded the title compounds in the crude state. Chromatographic separation of the resulting residue using (EtOAc-'petrol' 1:1) gave crystalline oxides 16 α , 16 β in a combined yield of 4.22 g (63%), mp 143–145 °C; $[a]_D^{21}$ +55.5 (c 4.0, CHCl₃).

For *isomer* **16a**, IR (KBr) $v_{\rm max}$ 1751, 1214 cm⁻¹; $\delta_{\rm H}$ 1.81–1.90 (1H, m, $J_{\rm P-H}$ 15.2, H^{ax}-5), 2.02 (3H, s), 2.05 (3H, s), 2.11 (3H, s), 2.17 (3H, s), 2.25–2.38 (1H, m, $J_{\rm P-H}$ 15.2, H^{eq}-5), 4.04–4.21 (2H, m), 4.37–4.57 (4H, m), 5.26 (1H, t, J 4.6), 5.30 (1H, dd, J 7.9, 3.3), 5.37 (1H, appt d, J 3.3), 5.51 (1H, appt d, J 3.3), 5.95 (1H, appt d, $J_{1',2'}$ 3.3 H-1'); $\delta_{\rm C}$ 20.46, 20.52 (2C), 20.64, 25.69 (d, $J_{\rm C-P}$ 7.27 C-5), 61.24, 66.80 (d, $J_{\rm C-P}$ 7.79, C-4), 67.21 (C-6), 68.23 (2C), 69.00 (2C), 94.09 (d, $J_{\rm C-P}$ 4.67, C-1'), 170.02 (2C), 170.05, 170.32; $\delta_{\rm P}$ -10.55 (Found: C, 43.45; H, 5.32; P, 6.88. $C_{17}H_{25}O_{13}P$ requires C, 43.60; H, 5.38; P, 6.61%).

For isomer **16β**, $\delta_{\rm H}$ 5.22 (1H, dd, $J_{1,2}$ 5.3, 3.3, H-1'); $\delta_{\rm C}$ 96.56 (d, $J_{\rm C-P}$ 4.4, C-1'); $\delta_{\rm P}$ -10.90.

4-Methylumbelliferyl 2,3,4,6-tetra-O-acetyl- β -D-galactopyranoside 17 6d

Propane-1,3-diyl 2,3,4,6-tetra-O-acetyl-β-D-galactopyranosyl phosphate **16α** (3 g, 6.4 mmol) was dissolved in CH_2Cl_2 (30 ml), and the solution was cooled to -78 °C and treated with TMSOTf (0.35 g, 1.6 mmol). To the resultant mixture was added 7-hydroxy-4-methylcoumarin (1.13 g, 6.41 mmol) whilst the temperature was maintained at -78 °C, and the mixture was stirred for an additional 0.5 h. Following this the reaction mixture was warmed to 0 °C, neutralised with aq. NaHCO₃ (30 ml), and extracted into CH_2Cl_2 (20 ml), and the extract was dried over Na_2CO_3 and concentrated *in vacuo*. The resultant

residue was purified by column chromatography (EtOAcfetrol' 1:1) to afford 17 (2.25 g, 68%), mp 142–143 °C; $[a]_{\rm D}^{21}$ -10.0 (c 1.3, CHCl₃); IR (KBr) $\nu_{\rm max}$ 1750, 1614; $\delta_{\rm H}$ 2.03 (3H, s), 2.09 (3H, s), 2.11 (3H, s), 2.20 (3H, s), 2.42 (3H, d, J 1.3), 4.08–4.24 (3H, m), 5.16 (1H, d, J 6.0), 5.17 (1H, dd, J 10.6, 4.0), 5.49 (1H, d, J 3.3), 5.51 (1H, dd, J 10.6, 7.3), 6.19 (1H, d, J 1.3), 6.93 (1H, dd, J 8.6, 2.6), 6.99 (1H, d, J 2.6), 7.52 (1H, d, J 8.6); $\delta_{\rm C}$ 18.48, 20.39, 20.46 (2C), 20.50, 61.31, 66.72, 68.26, 70.53, 71.32, 98.72 ($J_{\rm C-H}$ 160.18), 103.86, 113.00, 113.72, 115.34, 125.57, 152.08, 154.69, 159.15, 160.60, 169.21, 169.89, 170.04, 170.30; m/z (CI, NH₃) [Found: (M + Na), 529.1325. $C_{\rm 24}H_{\rm 26}{\rm NaO}_{\rm 12}$ requires m/z, 529.1322].

4-Methylumbelliferyl β-D-galactopyranoside 18^{6d}

Compound 17 (2 g, 3.95 mmol) was *O*-deacetylated as described for the preparation of 13 above. Recrystallisation from ethanol gave the title compound 18 as a white solid (1.17 g, 83%), mp 261–263 °C; IR (KBr) $v_{\rm max}$ 3521, 1721, 1614 cm⁻¹; $\delta_{\rm H}$ 2.38 (3H, s), 3.52–3.72 (2H, m), 4.54 (1H, d, *J* 4.6), 4.66 (1H, t, *J* 5.3), 4.90 (1H, d, *J* 6.0), 4.97 (1H, d, *J* 7.9), 5.24 (1H, d, *J* 5.3), 6.21 (1H, s), 6.99–7.02 (2H, m), 7.65 (1H, d, *J* 9.2); $\delta_{\rm C}$ 18.11, 60.44, 68.12, 70.15, 73.24, 75.74, 100.62, 103.17, 111.67, 113.46, 114.03, 126.41, 153.37, 154.44, 160.17, 160.29; m/z (CI, NH₃) [Found: (M + H), 339.1080. $C_{16}H_{19}O_8$ requires m/z, 339.1080].

4-Methylumbelliferyl 6-O-(tert-butyldiphenylsilyl)- β -D-galactopyranoside 19

4-Methylumbelliferyl β-D-galactopyranoside **18** (1 g, 2.9 mmol) was dissolved in DMF (10 ml) and the solution was cooled to 0 °C. To the resultant solution was added, with stirring, tertbutyldiphenylsilyl chloride (0.8 g, 2.9 mmol) and imidazole (0.4 g, 5.9 mmol) over a period of 5 min. Stirring was continued for 8 h at RT. The solvent was removed in vacuo, the residue was dissolved in dichloromethane (50 ml) and this solution was washed with water (20 ml). The organic layer was dried (Na₂SO₄), and the solvent removed in vacuo. Column chromatography (EtOAc-'petrol', 5:1) gave **19** (1.34 g, 79%), $[a]_D^{22}$ -30.4 (c 3.0, CHCl₃); IR (film) v_{max} 3413, 1727, 1614 cm⁻¹; δ_{H} 1.03 (9H, s), 2.30 (3H, s), 3.67–3.76 (2H, m), 3.90 (1H, d, J 6.0), 3.94 (2H, s), 4.01 (1H, dd, J7.9, 3.3), 4.91 (1H, d, J7.9), 6.09 (1H, d, J 1.3), 6.84 (1H, d, J 2.6), 6.95 (1H, dd, J 8.6, 2.6), 7.24–7.37 (7H, m), 7.60–7.65 (3H, m), 7.99 (1H, s); δ_C 18.48, 19.02, 26.67 (2C), 31.40, 63.13, 68.76, 71.10, 73.67, 75.21, 100.56 (J_{C-H} 160.43), 104.24, 112.52, 113.42, 114.77, 125.20, 127.67 (3C), 129.73, 132.76, 132.89, 135.45 (2C), 135.50 (2C), 152.36, 159.83, 160.96, 162.65; m/z (CI, NH₃) [Found: (M + H), 577.2263. C₃₂H₃₇O₈Si requires *m/z*, 577.2257].

4-Methylumbelliferyl 6'-*O*-(*tert*-butyldiphenylsilyl)-3'-*O*-(2",3", 4"-tri-*O*-acetyl-α-L-fucopyranosyl)-β-D-galactopyranoside 20

To a solution of 2,3,4-tri-O-acetyl-α,β-L-fucopyranosyl propane-1,3-diyl phosphate 6/7 (1 g, 2.4 mmol) in dichloromethane (10 ml) at -78 °C was added TMSOTf (0.54 g, 2,44 mmol). After 2 min, a solution of compound 19 (1.40 g, 2.44 mmol) in dichloromethane (10 ml) was added to the reaction flask. The reaction mixture was stirred at -78 °C for 1 h and was then allowed to warm up to 0 °C before quenching with aq. NaHCO₃ (10 ml). The organic layer was dried over (Na₂SO₄), and concentrated in vacuo. Column chromatography (EtOAc -'petrol', 6:4) gave 20 as a white crystalline solid (1.01 g, 52%), mp 109–111 °C; $[a]_{\rm D}^{22}$ –38.7 (c 3.0, CHCl₃); IR (KBr) $v_{\rm max}$ 3457, 1747, 1614 cm⁻¹; $\delta_{\rm H}$ 1.05 (9H, s), 1.17 (3H, d, J 6.6), 1.98 (3H, s), 2.04 (3H, s), 2.19 (3H, s), 2.37 (3H, s), 3.70 (1H, dd, J 5.9, 3.3), 3.90–3.96 (3H, m), 4.06–4.16 (3H, m), 4.70 (1H, d, J_{1,2} 7.3, H-1'), 5.02 (1H, br d, J 7.0), 5.08 (1H, d, $J_{1,,2}$ 3.3, H-1"), 5.13– 5.22 (2H, m), 5.27 (1H, dt, J 7.3, 2.6), 6.17 (1H, s), 6.94 (1H, d, J 2.0), 6.97 (1H, dd, J 8.6, 2.0), 7.28–7.44 (7H, m), 7.62–7.67 (4H, m); $\delta_{\rm C}$ 14.04, 15.78, 18.50, 19.04, 20.29, 20.42, 20.47, 20.51, 26.67, 60.29, 62.90, 67.70, 68.59, 69.63, 70.89, 71.87, 75.05, 80.81, 98.79 (C-1"), 101.63 (C-1'), 104.13, 112.95, 113.17, 115.01, 125.59, 127.58, 127.65 (2C), 127.69, 129.72, 129.74, 132.78, 132.95, 134.73, 135.46, 135.52, 152.02, 159.46, 160.73, 169.44, 169.97, 170.43; m/z (CI, NH₃) [Found: (M + NH₄), 866.3480. $C_{44}H_{56}NO_{15}Si$ requires m/z, 866.3419].

4-Methylumbelliferyl 3'-O-(2'',3'',4''-tri-O-acetyl- α -L-fucopyranosyl)- β -D-galactopyranoside 21

To a THF solution (5 ml) of silyl compound 20 (0.5 g, 0.58 mmol) was added tetrabutylammonium fluoride (0.1 M solution in THF; 15 ml) and the mixture was stirred for 12 h at RT. The solvent was removed in vacuo, the residue was dissolved in CH₂Cl₂ (10 ml), and the solution was washed with water (10 ml), dried over Na₂SO₄, and concentrated. The resultant material was purified by column chromatography (EtOAc-'petrol' 3:2) and gave thiol **21** (250 mg, 69%), $[a]_D^{23}$ -28.4 (c 1.8, CHCl₃); IR (film) v_{max} 3437, 1745, 1612 cm⁻¹; δ_{H} 1.26 (3H, d, J 7.3), 1.98 (3H, s), 2.04 (3H, s), 2.19 (3H, s), 2.40 (3H, s), 3.72– 3.78 (2H, m), 3.86–4.03 (2H, m), 4.08 (1H, d, J 7.3), 4.13 (1H, dd, J 7.3, 2.6), 4.28 (1H, dd, J 7.3, 6.6), 4.71 (1H, d, $J_{1',2'}$ 7.9, H-1'), 5.05–5.10 (2H, m), 5.14 (1H, appt d, J 7.9), 5.27 (1H, d, $J_{1'',2''}$ 2.6, H-1"), 6.18 (1H, s), 6.95–7.00 (2H, m), 7.51 (1H, d, J 9.9); $\delta_{\rm C}$ 15.84, 18.56, 20.30, 20.46, 20.55, 62.17, 67.90, 68.32, 68.65, 69.72, 69.85, 71.72, 74.76, 80.63, 98.86 (J_{C-H} 167.70, C-1"), 101.64 (*J*_{C-H} 161.99, C-1'), 104.10, 113.04, 113.23, 115.16, 125.70, 152.17, 154.84, 159.45, 160.81, 169.45, 170.02, 170.46; m/z (CI, NH₃) [Found: (M + NH₄), 628.1928. C₂₈H₃₈NO₁₅ requires m/z, 628.1928].

4-Methylumbelliferyl 1-O-(α -L-fucopyranosyl)- β -D-galactopyranoside 22

The disaccharide **21** (0.2 g, 0.32 mmol) was *O*-deacetylated as described for the preparation of compound **13** above. Recrystallisation from EtOH afforded the title compound **22** as a white solid (138 mg, 87%), mp 287–289 °C; [a]_D²¹ – 27.3 (c 0.2, H₂O); IR (KBr) $\nu_{\rm max}$ 3440, 1698, 1619 cm⁻¹; $\lambda_{\rm max}$ (H₂O) 320, 292, 252 nm; $\delta_{\rm H}$ (DMSO-d₆) 1.06 (3H, d, J 6.6), 2.40 (3H, s), 3.53–3.59 (2H, m), 3.63 (1H, dd, J 9.9, 3.3), 3.71 (1H, m), 3.78 (1H, dd, J 9.9, 7.3), 4.40 (1H, d, J 7.3), 4.45 (1H, q, J 3.3), 4.51 (1H, d, J 1.3), 4.69–4.72 (2H, m), 4.94 (1H, d, J 4.0), 5.08 (1H, d, J 6.6), 6.24 (1H, s), 7.00–7.10 (2H, m), 7.67 (1H, d, J 8.6); $\delta_{\rm C}$ (DMSO-d₆) 16.40, 18.08, 60.34, 67.33, 70.21 (2C), 70.97, 71.49, 73.32, 75.57, 78.31, 100.08 ($J_{\rm CH}$ 173.7), 103.04 ($J_{\rm CH}$ 159.4), 103.96, 111.71, 113.96, 114.27, 126.23, 153.31, 154.24, 160.07, 160.25; m/z (CI, NH₃) [Found: (M + NH₄), 502.1920. C₂₂H₃₂NO₁₂ requires m/z, 502.1924].

4-Methylumbelliferyl 6'-O-(tert-butyldiphenylsilyl)-2'-O-(2'',3'',4''-tri-O-acetyl- α -L-fucopyranosyl)- β -D-galactopyranoside

To a solution of the galactopyranoside **19** (1.01 g, 1.74 mmol) in CH_2Cl_2 (10 ml), containing $Et_4N^+Br^-$ (0.36 g, 1.74 mmol) and molecular sieves (4 g), was added 2,3,4-tri-O-acetyl- α -L-fucopyranosyl bromide **3** (0.62 g, 1.74 mmol). The resulting mixture was stirred at RT for 48 h. The solid residue was removed by passage through a pad of Celite and the filtrate was washed with water (10 ml). The CH_2Cl_2 layer was dried over

Na₂SO₄, and evaporated in vacuo. Column chromatography (EtOAc-'petrol' 1:4) gave compound 23 as a major (750 mg, 51%) and compound 20 as a minor product (390 mg, 27%), mp 79–81 °C; $[a]_{D}^{20}$ –27.32 (c 1.8, CHCl₃); IR (KBr) v_{max} 3452, 1745, 1614 cm^{-1} ; $\delta_{\text{H}} 1.05 \text{ (9H, s)}$, 1.16 (3H, d, J 6.6), 2.06 (3H, s), 2.17(6H, s), 2.37 (3H, s), 3.59 (1H, dd, J 4.0, 2.6), 3.68–3.72 (1H, m), 3.90–3.99 (2H, m), 4.10–4.22 (3H, m), 5.04 (1H, d, J 7.26, H-1'), 5.17–5.22 (2H, m), 5.27 (1H, dd, J7.3, 2.6), 5.46 (1H, dd, J 4.0, 3.3, H-1"), 6.16 (1H, s), 6.93 (1H, d, J 1.98), 6.97 (1H, dd, J 8.5, 2.6), 7.28–7.46 (7H, m), 7.62–7.65 (4H, m); $\delta_{\rm C}$ 14.09, 15.87, 18.53, 19.07, 20.31, 20.50, 20.54, 20.72, 20.93, 26.70, 60.32, 62.92, 67.74, 68.44, 69.67, 70.79, 71.91, 80.83, 98.83 (C-1"), 101.67 (C-1'), 104.22, 112.98, 113.22, 115.05, 125.61, 127.68 (2C), 127.73, 127.75, 127.81, 129.75, 129.77, 132.82, 135.47, 135.49, 135.55, 152.05, 152.23, 154.75, 159.50, 169.01, 170.01, 170.42; m/z (CI, NH₃) [Found: (M + NH₄), 866.5243. $C_{44}H_{56}NO_{15}Si$ requires m/z, 866.3075].

Acknowledgements

We thank the EPSRC for access to the mass spectrometry service at the University of Wales, Swansea (Director, Prof. D. E. Games).

References

- 1 (a) R. U. Lemieux, D. R. Bundle and D. A. Baker, J. Am. Chem. Soc., 1975, 97, 4076; (b) W. M. Watkins, in New Comprehensive Biochemistry, ed. J. Montreuil, H. Schachter and J. F. G. Vilegenhart, Elsevier, Amsterdam, 1995, vol. 29a, p. 313; (c) S. Canevari, D. Colombo, F. Compostella, L. Panza, F. Ronchetti, G. Russo and L. Toma, Tetrahedron, 1999, 55, 1469.
- 2 (a) H. Egge, A. Dell and H. von Nicolai, Arch. Biochem. Biophys, 1983, 224, 235; (b) T. Takamura, T. Chiba and S. Tejima, Chem. Pharm. Bull., 1981, 29 1076; (c) K. L. Matta, S. S. Rana, C. F. Piskorz and S. A. Abbas, Carbohydr. Res., 1984, 131, 247; (d) H. H. Lee, J. F. Schwartz, J. F. Harris, J. P. Carver and J. J. Krepinsky, Can. J. Chem., 1986, 64, 1912; (e) D. M. Whitfield, C. Ruzicka, J. P. Carver and J. J. Krepinsky, Can. J. Chem., 1987, 65, 693.
- 3 (a) J. B. Lowe, L. M. Stoolman, R. P. Nair, R. D. Larsen, T. L. Berhend and R. M. Marks, *Cell*, 1990, **63**, 475; (b) M. L. Phillips, E. Nudelman, F. C. A. Gaeta, M. Perez, A. Singhal, S.-I. Hakomori and J. C. Paulson, *Science*, 1990, **250**, 1130; (c) C. H. Wong, F. Moris-Varas, S.-C. Hung, T. G. Marron, C. -C. Lin, K. W. Gong and G. Weitz-Schmidt, *J. Am. Chem. Soc.*, 1997, **119**, 8152.
- 4 (a) G. Singh, I. Tranoy and H. Vankayalapati, *Chem. Commun.*, 1998, 2129; (b) G. Singh and H. Vankayalapati, *Tetrahedron Lett.*, 1999, **40**, 3925; (c) S. Hashimoto, T. Honda and S. Ikegami, *J. Chem. Soc.*, *Chem. Commun.*, 1989, 685; (d) H. Vankayalapati and G. Singh, *Tetrahedron: Asymmetry*, 2000, **11**, 125 and references therein.
- 5 (a) P. B. Høj, E. B. Rodriguez, R. V. Stick and B. A. Stone, J. Biol. Chem., 1989, 264, 4939; (b) D. H. Leaback and P. G. Walker, Biochem. J., 1961, 78, 151.
- 6 (a) R. Strachan, J. Wood and R. Hirschmann, J. Org. Chem., 1962,
 27, 1074; (b) D. Robinson, Comp. Biochem. Physiol., 1964, 12, 95;
 (c) D. Dunstan and L. Hough, Carbohydr. Res., 1972, 23, 425;
 (d) M.-C. Courtin-Duchateau and A. Veyrières, Carbohydr. Res., 1978, 65, 23.
- 7 R. Baker, H. A. Nunez, J. V. O'Connor and P. R. Rosevar, Can. J. Chem., 1981, 59, 2086.
- (a) G. J. F. Chittenden, *Carbohydr. Res.*, 1988, 183, 140; (b) M. L. Wolfrom and A. Thomson, *Methods Carbohydr. Chem.*, 1963, 2, 211.
 K. Bock and C. Pedersen, *J. Chem. Soc.*, *Perkin Trans.* 2, 1974, 293.
- 10 R. U. Lemieux and H. Driguez, J. Am. Chem. Soc., 1975, 97, 4069.
- 11 J. Fiandor, M. T. García-López, F. G. De Las Heras and P. P. Méndez-Castrillón, Synthesis, 1985, 1121.